NOM characterization and removal at six Southern African water treatment plants

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Abstract

Organic pollution is a major concern during drinking water treatment. Major challenges attributed to organic pollution include the proliferation of pathogenic micro-organisms, prevalence of toxic and physiologically disruptive organic micropollutants, and quality deterioration in water distribution systems. A major component of organic pollution is natural organic matter (NOM). The operational mechanisms of most unit processes are well understood. However, their interaction with NOM is still the subject of scientific research. This paper takes the form of a metastudy to capture some of the experiences with NOM monitoring and analysis at a number of Southern African Water Treatment Plants. It is written from the perspective of practical process selection, to try and coax some pointers from the available data for the design of more detailed pilot work. NOM was tracked at six water treatment plants using dissolved organic carbon (DOC) measurements. Fractionation of the DOC based on biodegradability and molecular weight distribution was done at a water treatment plant in Namibia. A third fractionation technique using ion exchange resins was used to assess the impact of ozonation on DOC. DOC measurements alone did not give much insight into NOM evolution through the treatment train. The more detailed characterization techniques showed that different unit processes preferentially remove different NOM fractions. Therefore these techniques provide better information for process design and optimisation than the DOC measurement which is routinely done during full scale operation at these water treatment plants. Further work will focus on streamlining and improving the reproducibility of selected fractionation techniques, characterization of NOM from different water sources, and synthesis of the results into a systematic, practical guideline for process design and optimisation.
1 Introduction

In most parts of the world there is increased concern towards the effects of organic pollution on drinking water treatment. Major challenges attributed to organic pollution include the proliferation of pathogenic micro-organisms, prevalence of toxic and physiologically disruptive organic micro-pollutants, and quality deterioration in water distribution systems. However, since the number of organic compounds present in water is large, it is very difficult to monitor them individually during full scale operation. Thus, surrogate measurements are widely used. Table 1 shows selected organic pollutants that are regulated by the USEPA and in South Africa. DOC is the organic matter passing through a 0.45 µm filter while the other parameters are classified as organic micropollutants. Evidently, organic micropollutants constitute a very small component of the organic pollution. The major fraction of the organic pollution is attributed to natural organic matter (NOM). NOM is a heterogeneous mixture of undefined structurally complex organic compounds derived from plants, animals and micro-organisms and their waste and metabolic products. Therefore NOM inevitably occurs in all natural water sources.

NOM had been implicated as the direct cause for the formation and proliferation of some organic micro-pollutants. Chlorination of NOM can lead to the formation of trihalomethanes (THMs), and reaction with ozone can form toxic peroxide radicals, and physiologically disruptive aldehydes among other harmful by-products. Additionally, NOM had been shown to be directly responsible for several physical and bio-chemical water quality problems. NOM could be responsible for the colour, undesirable taste and odour of natural waters; is a source of nutrients for heterotrophic bacteria; promotes bacterial re-growth and deposition of particles in the distribution system which compromises water quality and increases turbidity at the consumer. Furthermore, NOM interferes with the performance of several unit processes. NOM could be responsible for the high coagulant demand; rapid clogging of filters by biofilm growth on media; rapid saturation of activated carbon beds thereby increasing the regeneration frequency; high
disinfectant demand; inhibition of the impact of disinfectants; rapid decay of ozone; inhibition of precipitative processes which form the backbone of drinking water treatment; is a major membrane foulant and may inhibit the removal of organic micro-pollutants by activated carbon. Given the inevitable occurrence of NOM in all natural water sources, more attention should be directed towards the systematic consideration of NOM – its structure and occurrence in raw water sources, its flow and fate during water treatment processes, and its analytical fractionation into parts that can guide engineers towards more optimal process selection and design.

Larger treatment plants in Southern Africa rely almost exclusively on surface water sources, which are often compromised due to high return flows and indirect reuse. Water temperature is high, many surface water impoundments are eutrophic and NOM concentrations are high. Considering the often poor performance of wastewater treatment plants, changes in land use patterns due to rapid industrial development, increased use of chemicals in agriculture, and new settlements, good drinking water has to be produced from ever changing and compromised raw water sources. The water sector is understandably concerned about the levels of NOM in drinking water, finding legal expression as a dissolved organic carbon (DOC) guideline limit of 5 mg/l and a maximum 10 mg/l. This forced attention to the flow and fate of NOM in treatment plants, led to a number of smaller studies at different locations in recent years.

This paper takes the form of a metastudy to capture some of the experiences with NOM monitoring and analysis at a number of Southern African Water Treatment Plants (WTPs). It is written from the perspective of practical process selection, to try and coax some pointers from the available data for the design of more detailed pilot work. The main objectives are to:

- Demonstrate that the normal routine monitoring of DOC does not bring much insight into process performance.
- Enumerate selected methods for fractionating the NOM and their possible relevance to different treatment processes.
– Evaluate the performance of treatment processes by tracking the removal and transformation of NOM through different treatment trains using three different fractionation techniques.

– Assess the practical value and relevance of the information that can be extracted from these fractionation techniques.

2 Tracking unfractionated NOM during full-scale treatment

The removal and transformation of NOM during full scale treatment was tracked using DOC measurements. DOC varies with the type of water from approximately 0.5 mg/l for groundwater and seawater to over 30 mg/l for coloured water from swamps (Thurman, 1985). From the perspective of evaluating NOM treatment, Chen et al. (2007) grouped unit processes into four classes:

– pretreatment (includes pre-oxidation using ozone or permanganate)

– conventional (includes enhanced coagulation, dissolved air flotation, settling and rapid sand filtration)

– advanced (includes granular activated carbon, oxidation preceding biofiltration and membrane filtration)

– disinfection (includes free chlorine, chloramination, or sequential chlorination)

In this paper we present results from six water treatment plants (WTPs) in Southern Africa, namely: Vaalkop (Magalies Water, South Africa), Vereeniging (Rand Water, South Africa), Balkfontein (Sedibeng Water, South Africa), Stilfontein (Midvaal Water, South Africa), Von Bach (NamWater, Namibia) and Goreangab (Windhoek City Council, Namibia). The configurations of the WTPs studied are shown in Table 2 according to the classification of Chen et al. (2007). Pretreatment is only applied at Goreangab.
Conventional treatment forms the core of the NOM treatment at the Vereeniging, Balkfontein and Von Bach WTPs. Advanced treatment is applied at Vaalkop (where ozonation precedes GAC filtration), Stilfontein (where ozonation precedes RSF) and Goreangab (a comprehensive treatment scheme with ozonation, BAC, GAC and ultrafiltration as advanced treatment steps).

2.1 Experimental

During full scale operation, DOC is only measured in the raw and final water. In order to evaluate each individual unit process, additional sampling and laboratory analyses were required to supplement the routine monitoring data. Samples were collected from the raw water, after each unit process and from the final treated water. Two litres of each sample were collected at each site, stored in the refrigerator at 4°C and analysed within 24 h. DOC was measured by quantifying the amount of carbon dioxide formed upon incineration of a water sample (after filtration through a 0.45 µm membrane filter) in a TOC analyzer. This sampling and analysis programme formed part of a post-graduate project at the Department of Chemical Technology at the University of Johannesburg and is fully documented elsewhere (Matsebula, 2009).

2.2 Results

Figure 1 shows the DOC removal at the six WTPs for each unit process. The DOC levels in the raw water are very high (>10 mg/l) at all the WTPs except Von Bach. Therefore DOC removal is an important operational objective at these WTPs. However, the DOC removal is poor with DOC values greater than the guideline value of 5 mg/l in the final treated water except at Von Bach and Goreangab where the DOC is reduced to 4.2 mg/l and 2.0 mg/l respectively in the treated water.

Table 3 shows the DOC removal for each individual unit process. EC, DAF, and ST are primarily phase separation methods and have been grouped together in Table 3 for analysis. Evidently, DOC removal is inconsistent for each individual process.
for different raw water sources hinting at strong differences in the NOM composition between the raw water sources. This demonstrates that the monitoring of DOC only brings limited insight into process performance and thus can not be used for design or process optimization. Therefore further DOC characterization is required to reveal the NOM characteristics responsible for these differences.

NOM is a collection of organic molecules with highly versatile chemical and physical structures. Organic compounds found in natural waters can be classified into six major structural groups, namely: humic substances, hydrophilic acids, carboxylic acids, carbohydrates, amino acids, and hydrocarbons. From this composition, we can deduce six properties of NOM relevant to water treatment, namely: (1) humicity, (2) bioavailability, (3) molecular weight distribution, (4) functional groups, (5) hydrophobicity/hydrophilicity, and (6) charge distribution. It is not a simple matter to measure these properties – such characterization of NOM only recently became available. Tracking these properties of NOM through the treatment train should reveal more information about process performance than the analysis presented in this section.

3 Fractionation of NOM into biodegradable and non-biodegradable fractions

This section deals with the bio-available fractionation of the NOM identified above, which classifies NOM based on its ability to support microbial growth. This is important in order to evaluate the biostability of water and disinfection demand in the distribution system and the performance of individual unit processes. Bio-available NOM had been previously quantified as assimilable organic carbon (AOC) and biodegradable organic carbon (BDOC).

AOC is the part of DOC that can be easily assimilated by bacteria and converted to cell mass and indicates bacterial re-growth potential in the distribution system. AOC is mainly related to NOM with molecular weight less than 1000 Dalton and corresponds to 16–38% of DOC and 4–9% of the colour (Hem and Efraimsen, 2001). BDOC is a measure of both the DOC that is mineralised and assimilated by bacteria and indicates
disinfection demand and disinfection by-product formation potential (DBFP). The quotient AOC/BDOC may be regarded as an indication of the relative biological stability of the biodegradable organic compounds present in drinking water (Van der Kooij, 1990). AOC and BDOC therefore provide complementary information. Based on the above, BDOC should be more suitable as a treatment process control parameter while AOC should be more suitable for distribution system water quality control. Very few studies in South Africa have been done using AOC, but more recently BDOC had been included in studies from Namibia.

### 3.1 Experimental

For the determination of BDOC, an inoculum is added to a fixed mass of sample, aerated and kept at ambient temperature. Daily measurements of DOC are taken until no further change in DOC is observed. This takes 5–7 days for most natural water sources. The BDOC is then calculated as the difference between the initial DOC and the final DOC. The procedure is optimised for inoculum size, incubation period and aeration.

During the study reported here, biologically active anthracite was taken from an operational rapid sand filter at a water treatment plant. The advantage of this culture is that it is not a mono culture under strict sterile conditions, but a well adapted heterogeneous culture. The anthracite was pre-washed 10 times with distilled water until no DOC in the distilled water was detected. The initial DOC of the water sample to be tested (DOC₀) was then determined. 300 ml of water sample was mixed with 100 g of anthracite. The contents were mixed gently and the DOCᵢ of the mixture determined. The difference DOCᵢ-DOC₀ must be less than 0.2 mg/l according to the assay. Air flow at a rate of 2–4 l/h was bubbled continuously through the water-anthracite mixture at ambient temperature for 5 days. Every 24 h a 20 ml sample was taken and filtered through a 0.45 µm filter to determine the DOC. The difference between the DOCᵢ and the minimum DOCᵢ min reached after 5 days was regarded as the BDOC. The DOCᵢ min was taken as the non-biodegradable fraction (NBDOC).
This sampling and analysis programme formed part of research project at the New Goreangab Water Reclamation Plant (NGWRP) in Windhoek and is documented elsewhere in more detail (Menge et al., 2009).

3.2 Results

The New Goreangab Water Reclamation Plant uses a blend of water from a dam and effluent from a wastewater treatment plant as the raw water source in a ratio which is very variable ranging from 1:20 to 1:12. The configuration of the plant consists of pre-ozonation (O$_3$), enhanced coagulation (EC), dissolved air flotation (DAF), followed by sand filtration (SF), ozonation (O$_3$), and biological activated carbon (BAC). After BAC water passes through granular activated carbon (GAC), ultrafiltration (UF), chlorination (CL), and stabilization (ST). Grab samples were collected after each treatment step for analysis. Four replicates were collected during each sampling period. Samples were collected during the period from October 2007 to November 2008. Average values for the DOC tracking after various treatment units for one sampling run are shown in Table 4.

The following observations can be made from these results:

1. NBDOC and BDOC are 70% and 30% respectively in the raw water. This is consistent with the composition of the raw water i.e. contains a significant proportion of effluent from a wastewater treatment plant.

2. DAF preferentially removes the NBDOC fraction and is largely unresponsive to the biodegradable part of the DOC.

3. RSF targets mostly the BDOC fraction. Haarhoff and Van Staden (2006) demonstrated the existence of an active biofilm in rapid sand filters. Biological activity in this biofilm might be responsible for the consumption of the BDOC fraction. This unit process reduces the BDOC fraction to only 20% of the total DOC.
4. $O_3$ increases the BDOC fraction by 34%. This is consistent with the common understanding that ozone breaks down larger, complex molecules to smaller biodegradable units.

5. BAC reduces the BDOC fraction by 34%, an equal magnitude to the increase in BDOC during ozonation. Therefore $O_3$ and BAC are complimentary processes.

There is 100% removal of BDOC in the effluent and the remaining 1.97 mg/l of DOC in the effluent is entirely NBDOC. Baghoth et al. (2008) tracked DOC through the urban water cycle and noted that the biodegradable fraction was readily removed during drinking water treatment whereas the non-biodegradable fraction would persist throughout the urban water cycle. This is consistent with the observations reported here.

4 Fractionation of NOM by liquid size exclusion chromatography (LSEC)

Liquid size exclusion chromatography (LSEC) is a technique which fractionates NOM based on molecular weight distribution (MWD). Size is an important characteristic in water treatment as diffusion coefficients and removal efficiencies are directly dependent on the size of the solute. NOM exhibits different characteristics depending on its MWD both in terms of treatability and potential water quality problems. The design, integration and performance of processes such as coagulation, dissolved air flotation, solid media and membrane filtration, sorption on granular activated carbon, disinfection and potential re-growth in the distribution system are strongly influenced by the MWD of the NOM (Tadanier et al., 2000).

This sampling and analysis programme formed part of a research project at the Goreangab water treatment plant and is documented elsewhere in more detail (Jacquemet et al., 2007).
4.1 Experimental

The LSEC consists of a liquid chromatography column coupled to on-line ultraviolet absorbance (UVA), Fluorescence and DOC detectors. The chromatography column separates NOM molecules based on molecular size so that the largest molecules are eluted first and the smallest molecules are eluted last in the column. This is because molecules that are smaller than the pore size can enter the particles and will thus have longer transit time than larger molecules as the larger molecules can not penetrate the pores very deeply. The elution time can then be plotted against the DOC on one axis, the UVA, and/or the Fluorescence on the other axis. Using the resultant plot, NOM can then be characterised into high or low molecular weight and humic or non-humic fractions.

Using a calibration based on potassium hydrogenophthalate standard, the different peaks of the chromatogram were integrated to define and evaluate the proportion of each organic fraction. The chromatographable DOC (CDOC) was measured as the sum of all chromatographic fractions identified; hence this is the hydrophilic part of NOM. The hydrophobic fraction (HDOC) was then determined as the difference between the total DOC and CDOC.

The first fraction (PS) was detected after 25–35 min and this comprises the largest molecules attributed to polysaccharides, proteins and colloids. The second peak was observed after 35–50 min and is attributed to aromatic organic molecules of high molecular weight such as humic substances and the corresponding building blocks (HS+BB). The third fraction, detected after 50–60 min, comprises the smallest molecules and is attributed to low molecular weight organic acids and neutrals (A+N).
4.2 Results

The proportions of these fractions in the sewage effluent were: PS<5%, \( \text{HS}+\text{BB}=60–65\% \) and \( \text{A}+\text{N}=15\% \). Considering the ratio between dam water and sewage effluent respectively, the blend exhibited a composition similar to the sewage effluent. The removal of the three fractions is shown in Table 5 after each unit process. The following observations can be made:

1. DAF removes the highest molecular weight fractions better, while the removal of the intermediate and lower molecular weight fractions is comparable.

2. SF targets the highest and lowest molecular weight fractions. Adsorption is probably responsible for the removal of the high molecular weight fractions. Biodegradation in the biofilm is probably responsible for the removal of the lowest molecular weight fractions because they are likely to be highly biodegradable.

3. When ozonation is optimized for NOM treatment, we would expect a decrease in PS and a corresponding increase in A+N and/or SH+BB to reflect the breaking down of high molecular weight fractions into lower molecular weight fractions during ozonation. The observed performance of \( \text{O}_3 \) process does not conform to this operational objective. This reveals that this unit process may require optimisation to improve its performance.

4. The observed performance of BAC cannot be readily explained. This may be related to the preceding ozonation step, which may be less than optimal.

5. Results for GAC are consistent with the literature that it targets mostly low to intermediate molecular weight fractions.

6. Ultrafiltration is observed to target the high molecular weight fractions. This is probably due to size rejection by the membranes.
5 Fractionation of NOM by ion exchange resins

NOM can be characterised into hydrophobic and hydrophilic fractions based on their affinity for water. The hydrophobic fraction has more affinity for water and is composed mainly of low molecular weight carbohydrates, proteins and amino acids. The hydrophobic fraction has less affinity for water thus more soluble in organic solvents and is composed mainly of humic and fulvic acids. The affinity for water by the hydrophilic species is attributed to their charged/polarized molecular structure which enables them to form hydrogen bonds with water thus more readily soluble in water than polar solvents. The hydrophobic species tend to be electrically neutral and non-polar, and thus readily soluble in neutral and non-polar solvents.

The hydrophobic and hydrophilic species can be further partitioned based on their ability to donate or accept a proton into acidic, basic, and neutral sub-groups. The acidic group is comprised mainly of carboxylic acids and phenols and it is the anionic character of this functional group that gives NOM its aqueous solubility, high affinity for metal complexation, and buffer capacity. The basic group is composed of amines and amides and is found in amino acids, poly peptides and aquatic humic substances. Because they are basic, they are readily adsorbed by sediments (Thurman, 1985). The most important neutral functional groups of NOM are hydroxyl, ether, ketone, aldehyde, ester and lactone. These functional groups are present in aquatic humic substances, carbohydrates, tannins, and ketoacids and because they contain oxygen, they are able to form bonds with water molecules when they dissolve.

5.1 Experimental

Ion exchange resins at pre-adjusted pH values were used to fractionate the NOM. Amberlite XAD resins were used to isolate the hydrophobic fractions of NOM. Dianion WA-10 and Dowex 88 resins were used after the XAD resins to isolate the hydrophilic fractions. This procedure was done at pre-adjusted pH using appropriate solvents to elute the adsorbed fractions. Six NOM fractions were obtained, namely: hydrophobic acids
(HpoA), hydrophobic bases (HpoB), hydrophobic neutrals (HpoN), hydrophilic acids (HpiA), hydrophilic bases (HpiB) and hydrophilic neutrals (HpiN).

This technique was used to investigate the treatability of NOM by ozonation at Vaalkop WTP. Ozone is a highly reactive oxidant. Ozone is consumed in two steps: the rapid primary reaction and the rather slow decay step. The rapid primary reaction is attributed to the reaction of ozone with unsaturated carbon bonds in NOM. The result is an increase in the biodegradability of NOM and decrease in molecular weight.

Two litre samples were collected before and after ozonation and analysed for the six fractions. The methodology of this fractionation technique is illustrated in Fig. 2. This sampling and analysis programme formed part of a post-graduate project at the Department of Chemical Technology at the University of Johannesburg and is fully documented elsewhere (Nkambule, 2009).

5.2 Results

Table 6 shows the composition of the water before ozonation. Other studies have shown that the hydrophilic fraction is more abundant in natural water (between 55–70%) than the hydrophobic fraction while the basic fraction only constitutes 2–3% (Thurman, 1985). Therefore these water samples before ozonation do not conform to the observations of earlier researchers probably because of the effects of unit processes preceding the ozonation.

Figure 3 shows the removal of the six NOM fractions at Vaalkop WTP by ozonation during four sampling periods.

Fractionation of NOM after ozonation based on hydrophobicity and hydrophilicity reveals that ozonation removes mostly the hydrophilic fraction. A parallel fractionation into basic, acidic and neutral fractions shows that the basic fraction is more amenable to removal by ozonation whereas the acidic and neutral fractions increase marginally. Coupling of these properties reveals that the HpoB, HpiB, and HpiA are removed while there is an increase in the HpoA and HpoN fractions. The impact on the HpiN fraction was inconsistent. Furthermore, the magnitude of the changes in these fractions due to
ozonation varied significantly during the sampling period which may point to problems with reproducibility which require further investigation.

6 Further work

The operational mechanisms of most unit processes are well understood. However, their interaction with NOM is not well understood. Water treatment process designers would ultimately like to have a firm, rational footing for selecting processes for NOM removal. In its most ideal form, one would determine the different NOM components in a raw water source, and then select those processes that will reduce the most dominant NOM components. In this respect, the results of the research projects reported earlier are encouraging. It was possible to show that the NOM fractions responded very differently to different unit processes. Furthermore, the observed impact of some unit processes relies heavily on the technique for NOM characterisation. A project is therefore already underway towards the following:

– Improving and streamlining the most promising NOM fractionation procedures

– Improving the reproducibility of the selected NOM fractionation techniques.

– Monitoring five different types of surface source water in South Africa to determine their dominant NOM components

– Testing each of these water types against different unit processes

– Synthesis of the results from the above steps into a systematic, practical guideline for the selection of the most appropriate unit processes for NOM removal and their optimal design.
Summary and conclusions

NOM was tracked at six WTPs using DOC measurements. Fractionation of the DOC based on biodegradability and molecular weight distribution was done at a WTP in Namibia. A third fractionation technique based on the hydrophilic/hydrophobic characteristics of NOM was used to assess the impact of ozonation on DOC.

DOC measurements alone did not give much insight into NOM evolution through the treatment train. The performance of individual processes appeared inconsistent when the raw water source was varied hinting at a strong influence of other important constituents of the NOM. Fractionation of NOM based on biodegradability was more descriptive as the transformation of the BDOC and NBDOC fractions could be related to the operational mechanisms of the individual unit processes. This technique was especially informative for ozonation, rapid sand filtration and biological activated carbon. BDOC fractionation may thus be the analytical technique of choice when oxidation and biofiltration processes have to be analysed more deeply.

The results reported in this paper on molecular weight fractionation showed that rapid sand filtration targets the low and high molecular weight fractions, GAC targets the low to intermediate molecular weight fractions, and UF targets the high molecular weight fraction. These results are consistent with the operational mechanisms of these unit processes and thus can be used for operational analysis and/or optimization of these processes.

Fractionation using ion exchange resins showed wide variations during different sampling runs, which may point to problems with reproducibility. The technique, however imperfect it may be at present, did show up large differences in the NOM response to ozonation, which may turn out to be a potentially powerful tool for better understanding of NOM transformation and removal.

Taken together, there can be no argument that NOM fractionation, beyond the usual DOC analysis, is an important prerequisite for better understanding NOM, and designing of water treatment plants for its optimal removal. The results presented in this
paper, gleaned from studies by different parties, clearly demonstrated the high variability of NOM at the Southern African Water Treatment Plants which treat typical surface water, and the very different responses of different NOM fractions to different unit processes.

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References


**Table 1.** Regulated organic chemicals.

<table>
<thead>
<tr>
<th>Determinant</th>
<th>Unit</th>
<th>EPA max limit (2009)**</th>
<th>South African max limit (2005)***</th>
</tr>
</thead>
<tbody>
<tr>
<td>Dissolved organic carbon (DOC)</td>
<td>mg/l</td>
<td>Not mentioned</td>
<td>10</td>
</tr>
<tr>
<td>Total Trihalomethanes (TTHMs)</td>
<td>mg/l</td>
<td>0.080</td>
<td>0.2</td>
</tr>
<tr>
<td>Polychlorinated biphenyls (PCBs)</td>
<td>mg/l</td>
<td>0.0005</td>
<td>Not mentioned</td>
</tr>
<tr>
<td>Pesticides (sum)</td>
<td>mg/l</td>
<td>0.0005*</td>
<td>Not mentioned</td>
</tr>
<tr>
<td>1,1,2-Trichloroethane</td>
<td>mg/l</td>
<td>0.005</td>
<td>Not mentioned</td>
</tr>
<tr>
<td>Vinyl chloride</td>
<td>mg/l</td>
<td>0.002</td>
<td>Not mentioned</td>
</tr>
<tr>
<td>Haloacetic acids (HAA5)</td>
<td>mg/l</td>
<td>0.060</td>
<td>Not mentioned</td>
</tr>
</tbody>
</table>

* Value is from European Union guidelines  
** Source: Environmental Protection Agency, USA  
*** Source: South African National Standard (SANS) 241 Drinking Water Specifications
Table 2. WTPs configurations.

<table>
<thead>
<tr>
<th>Plant</th>
<th>Pre-treatment</th>
<th>Conventional Treatment</th>
<th>Advanced Treatment</th>
<th>Disinfection</th>
</tr>
</thead>
<tbody>
<tr>
<td>Vaalkop</td>
<td>None</td>
<td>EC</td>
<td>O₃ + GAC</td>
<td>Chloramination</td>
</tr>
<tr>
<td>Vereeniging</td>
<td>None</td>
<td>EC + ST</td>
<td>None</td>
<td>Chloramination</td>
</tr>
<tr>
<td>Balkfontein</td>
<td>None</td>
<td>EC + ST + RSF</td>
<td>None</td>
<td>Free chlorine</td>
</tr>
<tr>
<td>Stilfontein</td>
<td>None</td>
<td>DAF + ST</td>
<td>O₃ + RSF</td>
<td>Free chlorine</td>
</tr>
<tr>
<td>Von Bach</td>
<td>None</td>
<td>ST + RSF</td>
<td>None</td>
<td>Free chlorine</td>
</tr>
<tr>
<td>Goreangab</td>
<td>Pre-ozonation</td>
<td>EC + DAF + RSF</td>
<td>O₃ + BAC + GAC + UF</td>
<td>Free chlorine</td>
</tr>
</tbody>
</table>

EC = Enhanced Coagulation
ST = Settling Tank
RSF = Rapid Sand Filtration
DAF = Dissolved Air Flotation
O₃ = Ozonation
GAC = Granular Activated Carbon
BAC = Biological Activated Carbon
UF = Ultrafiltration
Table 3. DOC removal per unit process at the six WTPs.

<table>
<thead>
<tr>
<th>Plant</th>
<th>Phase Separation</th>
<th>RSF</th>
<th>O$_3$</th>
<th>RSF</th>
<th>GAC</th>
<th>BAC</th>
<th>Total Removal</th>
</tr>
</thead>
<tbody>
<tr>
<td>Vaalkop</td>
<td>5%</td>
<td>5%</td>
<td></td>
<td>8%</td>
<td></td>
<td></td>
<td>29%</td>
</tr>
<tr>
<td>Vereeniging</td>
<td>18%</td>
<td></td>
<td>−2%</td>
<td></td>
<td></td>
<td></td>
<td>20%</td>
</tr>
<tr>
<td>Balkfontein</td>
<td>11%</td>
<td>6%</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>29%</td>
</tr>
<tr>
<td>Stilfontein</td>
<td>19%</td>
<td>12%</td>
<td>−1%</td>
<td></td>
<td></td>
<td></td>
<td>37%</td>
</tr>
<tr>
<td>Von Bach</td>
<td>18%</td>
<td>15%</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>26%</td>
</tr>
<tr>
<td>Goreangab</td>
<td>29%</td>
<td>16%</td>
<td>9%</td>
<td></td>
<td></td>
<td></td>
<td>32% 81%</td>
</tr>
</tbody>
</table>
Table 4. Tracking NOM at the New Goreangab Water Reclamation Plant.

<table>
<thead>
<tr>
<th>Stage</th>
<th>DOC (mg/l)</th>
<th>BDOC (mg/l)</th>
<th>NBDOC (mg/l)</th>
<th>DOC Removal (%)</th>
<th>BDOC Removal (%)</th>
<th>NBDOC Removal (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Raw water</td>
<td>10.81</td>
<td>3.20</td>
<td>7.61</td>
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<td>–</td>
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</tr>
<tr>
<td>DAF</td>
<td>7.66</td>
<td>2.77</td>
<td>4.88</td>
<td>29</td>
<td>13</td>
<td>36</td>
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<tr>
<td>RSF</td>
<td>6.45</td>
<td>1.27</td>
<td>5.17</td>
<td>11</td>
<td>47</td>
<td>24</td>
</tr>
<tr>
<td>O₃</td>
<td>5.89</td>
<td>2.37</td>
<td>3.51</td>
<td>5</td>
<td>–34</td>
<td>–6</td>
</tr>
<tr>
<td>BAC</td>
<td>3.95</td>
<td>1.29</td>
<td>2.66</td>
<td>18</td>
<td>34</td>
<td>11</td>
</tr>
<tr>
<td>Final water</td>
<td>1.97</td>
<td>0</td>
<td>1.97</td>
<td>18</td>
<td>40</td>
<td>9</td>
</tr>
<tr>
<td><strong>Total Removal (%)</strong></td>
<td><strong>82</strong></td>
<td><strong>100</strong></td>
<td><strong>74</strong></td>
<td></td>
<td></td>
<td></td>
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Table 5. Removal rates of the three molecular weight fractions of NOM.

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<th>Stage</th>
<th>Removal Rate (%)</th>
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<td>PS</td>
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<tr>
<td>DAF</td>
<td>39</td>
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<tr>
<td>SF</td>
<td>13</td>
</tr>
<tr>
<td>O₃</td>
<td>1</td>
</tr>
<tr>
<td>BAC</td>
<td>-11</td>
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<td>GAC</td>
<td>19</td>
</tr>
<tr>
<td>UF</td>
<td>19</td>
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Table 6. Composition of the water before ozonation in mg/l.

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<th>Sampling Date</th>
<th>Hpo</th>
<th>Hpi</th>
<th>B</th>
<th>A</th>
<th>N</th>
<th>HpoB</th>
<th>HpoA</th>
<th>HpoN</th>
<th>HpiB</th>
<th>HpiA</th>
<th>HpiN</th>
</tr>
</thead>
<tbody>
<tr>
<td>Dec 2007</td>
<td>5.4</td>
<td>4.9</td>
<td>4.5</td>
<td>3.2</td>
<td>2.6</td>
<td>1.3</td>
<td>2.3</td>
<td>1.8</td>
<td>3.2</td>
<td>0.9</td>
<td>0.8</td>
</tr>
<tr>
<td>Jan 2008</td>
<td>5.7</td>
<td>5.6</td>
<td>4.6</td>
<td>3.5</td>
<td>3.2</td>
<td>1.5</td>
<td>2.5</td>
<td>1.7</td>
<td>3.1</td>
<td>1.0</td>
<td>1.5</td>
</tr>
<tr>
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<td>6.6</td>
<td>6.5</td>
<td>5.5</td>
<td>3.7</td>
<td>3.9</td>
<td>1.4</td>
<td>3.0</td>
<td>2.2</td>
<td>4.1</td>
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<td>1.7</td>
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<tr>
<td>Mar 2008</td>
<td>3.3</td>
<td>2.8</td>
<td>2.3</td>
<td>2.7</td>
<td>1.1</td>
<td>0.8</td>
<td>1.8</td>
<td>0.7</td>
<td>1.5</td>
<td>0.9</td>
<td>0.4</td>
</tr>
</tbody>
</table>
Fig. 1. DOC removal for the individual unit processes at the six water treatment plants.
Fig. 2. Steps applied during NOM fractionation using ion exchange resins.
Fig. 3. Removal of different NOM fractions by ozonation at Vaalkop WTP.